Optimization and Application of Endosomal Escape Vehicle (EEV™) Cell-Penetrating Peptides for Enhanced Delivery of Oligonucleotides and Genomic Medicines

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THERAPEUTICS

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OUR MISSION

To Treat Devastating Diseases with Intracellular Therapeutics





Entrada's pipeline includes a diverse array of high potential and high value assets; Each target disease has a substantial patient population with a significant unmet medical need





EEVTM PLATFORM DEVELOPMENT AND OPTIMIZATION

Endosomal Escape Vehicle (EEV™) Therapeutics

- Unique chemistry results in improved uptake and endosomal escape
- Cyclic structure designed to extend half life and increase stability
- Phospholipid binding potentially enables broad biodistribution to all cells

Mechanism of internalization conserved across species

Qian, Z. et al. ACS Chem. Biol. 2013; Qian, Z. et al. Biochemistry 2014; Qian, Z. et al. Biochemistry 2016; Sahni, A. et al. ACS Chem. Biol. 2020; Pei, D. Acc. Chem. Res. 2022.

Entrada seeks to solve a fundamental problem: a lack of efficient cellular uptake and escape from the endosome; Both are critical to intracellular target engagement and therapeutic benefit







Entrada has demonstrated intracellular uptake of unique moieties ranging from 1 kDa to 600 kDa

Anti	ibodies		Enzymes	5	Oligonuc	cleotides		Genomic Medicine		Antibody Drug Conjugates	
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300-3000 KDa	550-600 KDa	150 KDa	98 KDa	96 KDa	86 KDa	46 KDa	37 KDa	32 KDa	16 KDa	6 KDa	1-3 KDa
Genomic Medicine Formulation	Hybrid frataxin	Antibody	Thymidine phosphorylase	Purine nucleoside phosphorylase	Alanine- glyoxylate aminotransferase	Human frataxin	PTP1B catalytic domain	EGFP	Nanobody	Oligonucleotide	Various peptide cargos

DISCOVERY ENGINE FOR EEV THERAPEUTICS EEV-OLIGO EXAMPLE

Fit-for-purpose EEVs can be designed for target indications and modalities via iterative optimizations of EEV peptides through medicinal chemistry, *in vitro* and *in vivo* screenings



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DUCHENNE MUSCULAR DYSTROPHY: DELIVERY OF OLIGONUCLEOTIDES

DUCHENNE: SIGNIFICANT UNMET NEED





Duchenne is caused by mutations in the DMD gene, which lead to a lack of functional dystrophin,

causing progressive loss of muscle function throughout the body

Duchenne Franchise

ENTR-601-44

Phase 1: Positive preliminary data reported; Data to be presented October 2024

Phase 2: Regulatory filings expected Q4 2024

ENTR-601-45

Phase 2: Regulatory filings expected Q4 2024

ENTR-601-50

Phase 2: Regulatory filings expected 2025

Exon 51

Candidate selection expected in 2024

~40,000

people in the **US and Europe** have Duchenne¹

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EEV-PMO RESTORES MUSCLE INTEGRITY D2-mdx Mice



Robust exon 23 skipping after 4 monthly IV doses of EEV-PMO-23 in D2-*mdx* mice

Broad dystrophin expression and restoration of muscle integrity after four monthly IV doses of EEV-PMO-23 in D2-*mdx* mice





 D2-mdx mice (male, n=6-7) were treated with 4 monthly doses of either vehicle, 20 mg/kg unconjugated PMO-23 or 20 mg/kg PMO-23 equivalent of EEV-PMO-23, and the data were collected ~4 weeks after the last dose.

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SUPERIOR TO ALTERNATIVE PEPTIDES R6-PMO Example



EEV-PMO significantly improved exon 23 skipping after 3 days in *mdx* mice as compared to competitive R6-PMO



• EEV-PMO-23 demonstrates significantly improved PD effects after single 40 mg/kg IV dose in *mdx* mice

Values are shown as mean ± standard deviation; *p<0.05, **p<0.01; *mdx* is a DMD mouse model with a nonsense mutation in *DMD* exon 23; **PMO-23**, mouse *DMD* exon 23 skipping phosphorodiamidate morpholino oligomer; **IV**, intravenous.

CONSISTENT AND DURABLE EFFICACY OF EEV-PMO WAS DEMONSTRATED ACROSS SPECIES

Significant patient benefit is implied by data in the mouse and the monkey at clinically relevant levels; *in vitro* data suggests much higher target engagement in patient cells



- Single IV 60 mg/kg dose of ENTR-601-44
- Tibialis Anterior

- Post IV infusion of single 35 mg/kg dose of ENTR-601-44, robust exon 44 skipping observed in biceps of treated monkeys (n=3 per cohort) for at least 12 weeks
- Robust dose-dependent exon 44 skipping was observed in DMD patient-derived muscle cells harboring an exon 44 skip-amenable mutation

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hDMD transgenic mice (left) express full-length human dystrophin gene ('t Hoen, A.C. et al. *J. Biol. Chem.* 2008). **DMDA45** (right) are immortalized myoblasts from DMD patients harboring an out-of-frame exon 45 deletion and further differentiated into myotubes. Values are shown as mean ± standard deviation. **ENTR-601-44** is a DMD exon 44 skipping EEV-oligonucleotide **construct**. **DMD**, Duchenne muscular dystrophy; **hDMD**, human Duchenne muscular dystrophy; **IV**, intravenous.

ENTR-601-44 DATA SUMMARY



Significant patient benefit is implied by data in the mouse and the monkey at clinically relevant levels; in vitro data suggests much higher target engagement in patient cells

- ✓ High levels of exon skipping across *mdx*, D2-*mdx*, human dystrophin mouse and NHP studies
- Exon skipping translates to promising dystrophin production in heart and skeletal muscles
- ✓ Dystrophin production observed results in functional improvement in D2-*mdx* mouse
- Extended circulating half-life and durable exon skipping over 12+ weeks from a single injection of ENTR-601-44 was observed in the NHP

ENTR-601-44-101: Phase 1 clinical trial in healthy volunteers has completed dosing

- Cohorts 1, 2, and 3 have completed dosing and follow-up
- Fourth and final cohort has completed PK, PD, and safety assessments
- Preliminary results demonstrated exon 44 skipping with no treatmentrelated adverse events. Final results available by October 2024.
- Phase 1 clinical data will support the global clinical trial in patients*



PLATFORM EXPANSION: DELIVERY OF GENOMIC MEDICINES

EEV PLATFORM FOR GENOMIC MEDICINE DELIVERY



EEV-incorporated Lipid Nanoparticle (EEV-LNP) showed ~9-fold mRNA delivery improvement over LNP in HeLa cells, allowing the expansion of genomic medicine delivery



Hela cells were treated with 250 ng/mL EEV-LNP or LNP, both containing EGFP mRNA. Cells were harvested after 24 hours and analyzed for GFP expression by FACS analysis. One-way ANOVA was used for statistical comparison; *****p*<0.0001. **ANOVA**, analysis of variance; **EEV**, Endosomal Escape Vehicle; **EGFP**, enhanced green fluorescent protein; **FACS**, fluorescence-activated cell sorting; **EGFP**, enhanced green fluorescent protein; **LNP**, lipid nanoparticle; **mRNA**, messenger RNA.

IMPROVED mRNA DELIVERY: PRIMARY MACROPHAGES



EEV-LNP demonstrated dose-dependent and up to ~ 6-fold improvement in EGFP transfection efficacy in primary human macrophages *ex vivo* when compared to LNP control



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Primary CD14+ human monocytes were treated with granulocyte-macrophage colony-stimulating factor for 7 days to drive differentiation into macrophages. Cells were treated with EEV-LNP or LNP (both containing EGFP mRNA) and assessed EGFP expression by (A) fluorescence microscopy or (B) ELISA 24 hours post-treatment. Two-way ANOVA was used for statistical comparison; *****p*<0.0001. **ANOVA**, analysis of variance; **EEV**, Endosomal Escape Vehicle; **EGFP**, enhanced green fluorescent protein; **ELISA**, enzyme-linked immunosorbent assay; **LNP**, lipid nanoparticle; **mRNA**, messenger RNA.

GENE EDITING EFFICIENCY WITH EEV-LNP

EEV-LNP demonstrated dose-dependent gene editing, with editing efficiency consistently higher than LNP or transfection across a wide concentration range of Cas9 mRNA



EEV-LNP based co-delivery of Cas9 mRNA/gRNA showed dose-dependent gene editing *in vitro*

 Gene editing efficiency is quantified by FACS as percent of GFP-negative population in HEK293-uGFP cell line

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- EEV-LNP resulted in higher gene editing efficiency than LNP alone as well as transfection by MessengerMAX[™]
 - Dose dependent delivery of gene editing with EEV-LNP with an EC50 around 40 ng/mL
 - ~33-fold improvement at a dose as low as 40 ng/mL Cas9 compared to LNP

1. Li X, et al. *J Biol Chem.* 1998. After 48 hours of treatment, HEK293-uGFP6 cells were analyzed by flow cytometry to quantify the percentage of GFP-knockout cells. Two-way ANOVA test was used for statistical comparison; *****p*<0.0001. **ANOVA**, analysis of variance; **EC50**, half maximal effective concentration; **EEV**, Endosomal Escape Vehicle; **GFP**, green fluorescent protein; **gRNA**, guide RNA; **LNP**, lipid nanoparticle; **mRNA**, messenger RNA; **uGFP**, unstable green fluorescence protein.

GENE KNOCKDOWN WITH EEV-LNP COMPARED TO LNP ALONE

EEV-LNP showed 26-fold higher Cas9 expression at 5 hours post-transfection compared to LNP, demonstrating enhanced gene editing efficiency in primary macrophages

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Human primary macrophages were treated for 5 hours with EEV-LNP or LNP containing Cas9 mRNA and gRNA targeting B2M gene. (Left) Cas9 protein expression was quantified by Western blot at 5 and 24 hours post-treatment. (Right) B2M knockdown was assessed at 7 days post-treatment by FACS analysis. One-way ANOVA was used for statistical comparison; *****p*<0.0001, ***p*<0.001. **ANOVA**, analysis of variance; **AU**, arbitrary unit; **EEV**, Endosomal Escape Vehicle; **FACS**, fluorescence-activated cell sorting; **gRNA**, guide RNA; **LNP**, lipid nanoparticle; **mRNA**, messenger RNA.

EEV-LNP mRNA DELIVERY COMPARED TO ELECTROPORATION

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EEV-LNP demonstrated 5-fold improvement in mRNA delivery compared to electroporation, with reduced impact on cell viability

EEV-LNP vs. Electroporation (24 hours post-treatment)



Viability of Primary Macrophages



Green: Live cells (EGFP) Red: Necrotic/Dead Cells (Propidium Iodide)

EGFP: **p*<0.05 vs. untreated; *****p*<0.0001 vs. untreated and electroporation *Viability:* *****p*<0.0001 vs. untreated and EEV-LNP

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Human primary macrophages were differentiated from monocytes isolated from PBMCs and treated with EEV-LNP or electroporated (EGFP mRNA dose 2500 ng/10⁶ cells) using Lonza nucleofection method. EGFP expression was measured by ELISA after 24 hours (left). Cell viability quantified by LIVE/DEAD[™] Cell Imaging Kit (right). One-way ANOVA was used for statistical comparison; **ANOVA**, analysis of variance; **EEV**, Endosomal Escape Vehicle; **EGFP**, enhanced green fluorescent protein; **ELISA**, enzyme-linked immunosorbent assay; **LNP**, lipid nanoparticle; **PBMC**, peripheral blood mononuclear cell.

SIRPα KNOCKDOWN IN PRIMARY MACROPHAGES WITH EEV-LNP

SIRPa Knockdown in Primary Macrophages (FACS)

EEV-LNP demonstrated greater knockdown of SIRPα by FACS or NGS compared to LNP alone, with no significant impact on cell viability

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SIRPa Knockdown (NGS)



Human primary macrophages were differentiated from monocytes isolated from PBMCs with GM-CSF and treated with 1000 ng/ml EEV-LNP or LNP containing Cas9 mRNA and multiple gRNAs targeting SIRPα gene. Cell viability quantified by Sytox Blue™ Dead Cell Stain. SIRPα level was quantified using CD147. Percent Gene Editing was determined by NGS and analyzed by CRISPResso2. SIRPα is a membrane glycoprotein that inhibits macrophage function. CD80 is a co-stimulatory marker expressed on M1 macrophages. *p<0.05; **p<0.01; ****p<0.001; ****p<0.0001. One-way ANOVA was used for statistical comparison. **ANOVA**, analysis of variance; **EEV**, Endosomal Escape Vehicle; **FACS**, fluorescence activated cell sorting; **FITC**, fluorescencin isothiocyanate; **LNP**, lipid nanoparticle; **PBMC**, peripheral blood mononuclear cell; **FACS**, fluorescence-activated cell sorting; **NGS**, next-generation sequencing; **ns**, not significant; **SIRP**α, signal regulatory protein alpha.

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